

Enantioselective Synthesis of 3-Hydroxypiperidin-2-ones

Gary Gibbs, Martin J. Hateley, Lee McLaren, Matthew Welham and Christine L. Willis*

School of Chemistry, University of Bristol, Cantock's Close, Bristol BS8 1TS, UK

Received 11 November 1998; accepted 23 November 1998

Abstract: An efficient synthesis of (S)- and (R)-3-hydroxypiperidin-2-ones from methyl 5-nitro-2-oxopentanoate is described. A one-pot enzyme catalysed hydrolysis of the ester and reduction of the ketone gave enantiopure 2-hydroxy-5-nitropentanoic acids which on esterification, catalytic hydrogenation over a platinum(IV) oxide catalyst and intramolecular cyclisation gave the target compounds in 93% overall yield and >99% ee. © 1999 Elsevier Science Ltd. All rights reserved.

Hydroxylated five and six-membered ring nitrogen containing heterocycles are widespread in nature and are components of many biologically active compounds. Recently we described the synthesis of (S)- and (R)-3-hydroxypyrrolidin-2-ones (S)-1 and (R)-1, valuable building blocks for the preparation of nitrogen containing heterocycles.

Scheme 1

Our approach involved the lactate dehydrogenase (LDH) catalysed reduction of the sodium salt of 4-benzyloxycarbonyl(Z)-amino-2-oxobutanoic acid 5 (prepared from ethyl 3-Z-aminopropanoate 2 via a β -ketosulfoxide 3 according to a literature procedure³) followed by deprotection of the amine and cyclisation (Scheme 1). This was the first report of an LDH catalysed reduction of an α -keto acid containing a nitrogen functionality in the side chain. Use of commercially available LDH from Bacillus stearothermophilus (BS-LDH)⁴ for the reduction of 5 gave (S)-2-hydroxy acid (S)-6 in 91% yield while LDH from Staphylococcus epidermidis (SE-LDH)⁵ gave the enantiomer (R)-6 in 95% yield. In each case the cofactor NADH was recycled using the formate/formate dehydrogenase (FDH) method described by Shaked and Whitesides.⁶ We anticipated that we could extend the utility of this

$$Z_{H} \xrightarrow{CO_{2}Me} Z_{H} \xrightarrow{CO_{2}R} Z_{H} \xrightarrow{CO_{2}R} Z_{H} \xrightarrow{OH} Z_{H} \xrightarrow{OH} Z_{H} Z_{H} \xrightarrow{CO_{2}R} Z_{H} \xrightarrow{OH} Z_{H} Z$$

Scheme 2

approach to the synthesis of enantiomerically pure six membered ring homologues of γ -lactam 1, namely 3-hydroxypiperidin-2-one 7 via LDH catalysed reduction of Z-protected 5-amino-2-oxopentanoic acid 9 (Scheme 2).

First we examined the synthesis of the required α -keto ester 10 by an analogous procedure to that used for the preparation of 4 (Scheme 1). Although treatment of ethyl 4-Z-aminobutanoate 11 with dimsyl sodium gave β -keto sulfoxide 12, further attempted elaboration of 12 to α -keto ester 10 by reaction with N-bromosuccinimide produced only a complex mixture of products. Clearly an alternative approach for the synthesis of 10 was required. Wasserman and coworkers have described a valuable method for the homologation of a carboxylic acid to an α -keto ester via ozonolysis of a β -ketocyanophosphorane. Coupling 4-Z-aminobutanoic acid 13 with cyanomethylenetriphenylphosphorane in the presence of EDCI/DMAP gave 14 in a disappointing 35% yield along with pyrrolidinone 15 (Scheme 3). Ozonolysis of 14 in methanol-dichloromethane returned only the cyclised product 15, none of the required α -keto ester 10 was obtained. In contrast, methyl 4-Z-amino-2-oxobutanoate 4 was readily prepared from 3-Z-aminopropanoic acid 16 using this approach. Thus it was evident that the problems inherent to the synthesis of the required α -keto ester 10 originate from the nucleophilic nitrogen leading to cyclisation to the 5-membered ring lactam, whereas in the synthesis of methyl 4-Z-amino-2-oxobutanoate 4 cyclisation to the more strained β -lactam was not observed.

One way to circumvent these problems was to design an α -keto acid with a non-nucleophilic nitrogen containing functionality in the side-chain which, following the enzyme catalysed reduction of the ketone, may be converted to an amine to effect the required cyclisation to 3-hydroxypiperidin-2-one. A nitro group can be considered as a masked amine and so our new target α -keto ester was methyl 5-nitropentanoic acid 21. Use of a nitro group has two potential advantages over a Z-protected amine, first it is non-nucleophilic so cyclisations will not occur and secondly its compact size may allow an improved rate of turnover in the enzyme compared with a more bulky Z-protected amine (cf.

SE-LDH catalysed reduction of the sodium salt of Z-protected α -keto acid 5 took 7 days to reach completion on a 1 mmol scale giving a 95% yield of (R)- 6^2).

CCO₂tBu

i

$$O_2N$$
 O_2N
 O_2N

Reagents: i) MeNO₂, DBU; ii) TFA; iii) SOCl₂; iv) Ph₃PCHCN, BSA; v) O₃, MeOH, CH₂Cl₂; vi) BS-LDH, FDH, NADH; vii) H₂. PtO₂; viii) SE-LDH, FDH, NADH.

Scheme 4

Methyl 5-nitro-2-oxopentanoate 21 was prepared in five steps from commercially available starting materials (Scheme 4). Conjugate addition of nitromethane to tert-butyl acrylate in the presence of DBU8 gave a quantitative yield of methyl 4-nitrobutanoate 17. Removal of the tert-butyl group, conversion of the resulting carboxylic acid 18 to an acid chloride 19, and coupling with cyanomethylenetriphenylphosphorane using bis(trimethylsilyl)acetamide (BSA) as a proton scavenger furnished 20 in good yield. Ozonolysis of 20 in methanol-dichloromethane gave α -keto ester 21 in 51% yield over the five steps. It was then necessary to convert ester 21 into the corresponding α -keto acid. This may be achieved by saponification but a more convenient approach is to use a dual enzyme procedure to hydrolyse the ester and reduce the ketone in a one-pot process.⁹ Thus 21 was incubated with a lipase from Candida rugosa (CRL)10 prior to the addition of BS-LDH, FDH and the cofactor NADH. After work up, (S)-2-hydroxy-5-nitropentanoic acid was isolated and then methylated with diazomethane to give 2-hydroxy ester (S)-23 in 93% yield over the 3 steps. The (R)-enantiomer was obtained in an analogous manner but using SE-LDH to catalyse the reduction of keto acid 22. Interestingly on a 1 mmol scale the SE-LDH catalysed reduction of 22 was complete within 2 days which compared favourably with the 7 days required for the reduction of the more bulky 4-Z-amino-2oxobutanoic acid 5. ¹H- and ¹⁹F-NMR analysis of the (R)-(+)-MTPA (Mosher¹¹) derivatives of the hydroxy esters 23 were used to determine the enantiomeric purity of each product from the enzyme

catalysed reductions, and was found to be >99%ee in both cases. The chemical shift differences between the diastereomers were entirely consistent with the expected absolute configuration at C-2 according to correlation models of Mosher¹¹ and Yamaguchi. ¹²

Reduction of the nitro group in (S)-23 to the amine was accomplished by hydrogenation over a platinum(IV) oxide catalyst¹³ at atmospheric pressure, and spontaneous cyclisation gave (S)-3-hydroxypiperidin-2-one (S)-7 in quantitative yield. Spectroscopic data, melting point and optical rotation of this product were in complete agreement with those reported in the literature.¹⁴ (R)-3-Hydroxypiperidin-2-one (R)-7 was obtained in an analogous manner from (R)-23.

In conclusion, an efficient chemoenzymatic method has been developed for the enantioselective synthesis of (S)- and (R)-3-hydroxypiperidin-2-ones in 93% yield and >99%ee from 5-nitro-2-oxopentanoic acid 22.

Acknowledgements.

We are very grateful to the following for financial support: BBSRC (LM), EPSRC (GG and MJH) and Genzyme (MJH) and to Dr. Howard Marriage for valuable discussions.

References

- 1. For reviews see for example: Pinder, A. R., Nat. Prod. Rep., 1992, 9, 491; Michael, J. P., Nat. Prod. Rep., 1997, 14, 619.
- 2. Bentley, J. M.; Wadsworth, H. J. and Willis, C.L, J. Chem. Soc., Chem. Commun., 1995, 231.
- 3. Iriuchijima, S. and Ogawa, M., Synthesis, 1982, 41.
- Bur, D.; Luyten, M. A.; Wynn, H.; Provencher, L. R.; Jones, J. B.; Clarke, A. R. and Holbrook,
 J. J., J. Am. Chem. Soc., 1989, 67, 1065.
- 5. Kim, M.-J. and Kim, J. Y., J. Chem. Soc., Chem. Commun., 1991, 326.
- 6. Shaked, Z. and Whitesides, G. M., J. Am. Chem. Soc., 1980, 102, 7104.
- 7. Wasserman, H. H. and Ho, W.-B., J. Org. Chem., 1994, 59, 4364.
- 8. Bäckvall, J.-E.; Ericsson, A. M.; Plobeck, N. A. and Juntunen, S. K., Tetrahedron Lett., 1992, 33, 131.
- For a general procedure for the dual lipase/ oxidoreduction of an α-keto ester see: Macritchie, J. A.;
 Silcock, A. and Willis, C. L., Tetrahedron: Asymmetry, 1997, 8, 3895.
- 10. Bhalerao, U. T.; Dasaradhi, L.; Neelakantan, P. and Fadnavis, N. W., J. Chem. Soc., Chem. Commun., 1991, 1197.
- 11. Dale, J. A. and Mosher, H. S., J. Am. Chem. Soc., 1973, 95, 512.
- 12. Yasuhara, F. and Yamaguchi, S., Tetrahedron Lett., 1980, 21, 2827.
- 13. Secrist III, J. A. and M. W. Logue, J. Org. Chem., 1972, 37, 335.
- 14. Hua, D. H.; Zhang, F. and Chen, J., J. Org. Chem., 1994, 59, 5084; Hunter, A. and Woodward, H. E., Biochem. J., 1941, 35, 1298.